



Special Issue: Agricultural Productivity and Sustainability Improvement in Tropical Region

Potential of *Aloe vera* for Treatment of Infection with *Aeromonas hydrophila* Bacteria on Koi Fry

Rosidah Rosidah^{1*}, Angga Nugraha¹, Yuli Andriani¹, Kiki Haetami¹, Olga Anne² and Agus Heri Purnomo³

¹Department of Fisheries, Faculty of Fisheries and Marine Science, Universitas Padjadjaran, Jl. Raya Bandung, Sumedang Km 21, Jatinangor 45363, West Java, Indonesia; ²Technology and Natural Sciences Faculty, Klaipeda University, Herkaus Manto g. 84, Klaipeda 92294, Lithuania; ³Research Center for Marine and Fisheries Sosio-economics, Ministry of Marine Affairs and Fisheries, Republic of Indonesia, Gedung Mina Bahari 1, Jl. Pasir Putih 1, Ancol, Jakarta 14430, Indonesia.

Abstract | This research aims to study the ability of *Aloe vera* (Burm.f.), extract to treat Koi fry [*Cyprinus rubrofasciatus* (Lacépède, 1803)] which is attacked by *Aeromonas hydrophila* (Chester, 1901). The method used was the experimental method using a completely randomized design with five treatments and three replications was applied. The treatment used was immersion with *A. vera* extract with a concentration of treatment A (0 mg kg⁻¹), B (150 mg kg⁻¹), C (300 mg kg⁻¹), D (450 mg kg⁻¹) and E (600 mg kg⁻¹). The variables observed were *in vitro* test, LC₅₀-24-h test, clinical symptoms which included damage to fish body, fish feeding response and fish response to shock which were analyzed descriptively comparatively. The results showed *A. vera* extract has the ability as an antibacterial with inhibition zone diameter reaching 20.49 mm at a concentration of 5 000 mg kg⁻¹ and LC₅₀ test 48 h (*in vivo* test) of 1 157.16 mg kg⁻¹, so for the treatment of fish use concentrations below that concentration. The use of *aloe vera* extract with a concentration of 300 mg kg⁻¹ for 48 h was effective to treat koi fry infected with *A. hydrophila* by healing relatively fast body damage, response to feed and shock is normal and fast.

Received | March 03, 2021; **Accepted** | June 21, 2021; **Published** | October 27, 2021

***Correspondence** | Rosidah Rosidah, Department of Fisheries, Faculty of Fisheries and Marine Science, Universitas Padjadjaran, Jl. Raya Bandung, Sumedang Km 21, Jatinangor 45363, West Java, Indonesia; **Email:** ros_ahdi@yahoo.com

Citation | Rosidah, R., A. Nugraha, Y. Andriani, K. Haetami, O. Anne and A.H. Purnomo. 2021. Potential of *Aloe vera* for treatment of infection with *Aeromonas hydrophila* bacteria on koi fry. *Sarhad Journal of Agriculture*, 37(Special issue 1): 64-74.

DOI | <https://dx.doi.org/10.17582/journal.sja/2021/37.s1.64.74>

Keywords | Alternative medicine, Clinical symptoms, *Cyprinus rubrofasciatus*, Fish disease, Ornament fish

Introduction

One of the highly prospective ornamental fish, the technology that is controlled by farmers is Koi fry [*Cyprinus rubrofasciatus* (Lacépède, 1803)]. The marketing of Koi by exporters includes Germany, Africa, England, Singapore, and America. The constraints faced by exporters to meet consumer demand are the low availability of quality Koi fish (Triyanti and Yulisti, 2012), while the obstacles faced by farmers are the attack of Motile *Aeromonas Septicemia* (MAS) or red spot disease caused by

bacteria *Aeromonas hydrophila* (Chester, 1901). One of the diseases that often attack freshwater fish, one of which is Koi, especially in the fry stages.

Aeromonas hydrophila is an opportunistic type of pathogenic bacteria and can cause systemic diseases and result in mass death (Peatman *et al.*, 2018). *A. hydrophila* is a gram-negative, rod-shaped, size (0.8 to 1.0) µm × (1.0 to 3.5) µm. These bacteria can move (motile) and do not form spores. These bacteria are facultatively anaerobic, fermentative, positive cytochrome oxidase, positive catalase and produce H₂S

(Wassif, 2018). Laith and Najiah (2013) stated that the clinical symptom of fish attacked by *A. hydrophila* is inflammation with a characteristic swelling in the injection site. These symptoms continue with haemorrhagic (bleeding) which is characterized by the discharge of blood on the skin, the flake fin and slow swimming fins, exophthalmia (swollen eyes), ulcers (ulcers in the body) and necrosis are symptoms that are characterized by the appearance of damaged and decaying flesh.

Disease prevention in the cultivation system generally uses antibiotics. However, the use of antibiotics can cause a resistant effect on pathogenic bacteria and cause pollution to the environment (Kraemer *et al.*, 2019). Therefore, other alternative drugs that are more environmentally friendly and do not cause a resistant effect on bacteria, *A. vera* extract are needed.

According to Khan and Ayub (2020) *A. vera* leaves have a wide therapeutic effect, including being used as a diarrhea medicine, boosting the immune system, preventing tumors, maintaining the liver, preventing gastric inflammation, healing inflamed or injured tissue etc. Kar and Bera (2017) stated that, all parts of extracted *A. vera*, contain polysaccharides (acemannan and glucomannan), flavonoids, tannins, and saponins. According to Quezada *et al.* (2017), the two main polysaccharides contained in *A. vera* are glucomannan and acemannan. Glucomannan acts to replace skin tissue and reduce pain due to injury. Friendship can accelerate wound healing. Flavonoids are anti-inflammatory so can reduce inflammation and help reduce pain if there is bleeding or inflammation in the wound (Panche *et al.*, 2017). Tanin is useful as an antiseptic and also for the treatment of burns by prescribing proteins (Huang *et al.*, 2018). The mechanism of action of saponin as an antibacterial is to damage the constituent components of peptidoglycan on the bacterial cell wall (lipoprotein). This results in decreased lipid surface tension, changing cell permeability, and the mechanism can cause bacterial cell function to be abnormal, and lysis and dead cell bacteria (Hamouda and Baker, 2019).

The right way to treat Koi fish which is attacked by MAS is by dipping. According to (Kumar *et al.*, 2018), that treatment with an immersion system is the most applicable method compared to injection and feeding because it can simplify the treatment process especially for fish in the size of on a large scale.

Materials and Methods

This research includes preliminary research and main research. Preliminary research is carried out on March 4 to 10, 2019 and the main research will be conducted for 2 wk (weeks) in March and April. This research took place at the Gd 4 Aquaculture Laboratory, Fisheries and Agricultural Engineering and Biotechnology Laboratory; Gd. 3 Faculty of Fisheries and Marine Sciences, Padjadjaran University, Indonesia. The method used in this research is the experimental method using Completely Randomized Design (CRD) with five treatments and three replications. The treatment given was soaking Koi fry infected with *A. hydrophila* in *A. vera* extract with a concentration based on the results of *in vitro* and *in vivo* tests, as follows:

Treatment A: Control treatment without *A. vera* immersion; Treatment B: 150 mg kg⁻¹ *A. vera* extract; Treatment C: 300 mg kg⁻¹ *A. vera* extract; Treatment D: 450 mg kg⁻¹ *A. vera* extract; Treatment E: 600 mg kg⁻¹ *A. vera* extract.

Research procedure

The preparation of experimental Koi fry: The experimental fish used in this study originated from Center for Research and Development of Freshwater Aquaculture (BBPBAT- Balai Besar Perikanan Budidaya Air Tawar) in Sukabumi, West Java, sizing of 8 cm to 12 cm in total length, as much as 80 fishes for the initial study, 300 fishes for the further study. The experimental fishes were acclimatized for 7 d (days) to find out the healthy fish.

The preparation of aquarium: As much as 15 aquariums sizing of 40 cm × 30 cm × 20 cm were used in this study. The aquariums were cleaned, then the water was added fully and as much as 30 mg L⁻¹ of chlorine was added to sterilize the containers for 24. The aquariums were dried before being used. After that, the aquariums were filled with 20 L of water and were aerated.

The preparation of *A. vera* extract

A. vera extract was prepared by maceration method (Sopiah *et al.*, 2018). As much as 3 000 g of wet *A. vera* were dried for 14 d. The dried *A. vera* were performed by using 92 % methanol solution as much as 35 L for 3 d. The filtrate was filtered by using Whatman paper filter number 42. The maceration

filtrate was evaporated and concentrated by using rotatory evaporator at 80 °C to obtain the *A. vera* thick extract as much as 228.3 g.

Preliminary in vitro tests

The zone of inhibition used to test the effectiveness of *Aloe vera* extract as an antibacterial to inhibit the growth of *A. hydrophila* by using the [USEPA Method \(2002\)](#). The zone of inhibition was used disk diffusion test with four variances of *Aloe vera* extract concentration: 10 mg kg⁻¹, 100 mg kg⁻¹, 1 000 mg kg⁻¹, and 5 000 mg kg⁻¹ for 48 h. The material and equipment were sterilized using an autoclave. The disc paper was put to a petri dish with NA medium and 1 mL of *A. hydrophila* inoculation (the bacteria density was 10⁸ CFU mL⁻¹). The petri dish then was incubated for 24 h at 30 °C. The zone of inhibition was measured by using a caliper.

LC50-24-hour test

The LC50 test of *A. vera* extract was done to measure the short-term poisoning potential which causes 50 % of mortality. The concentration for LC50 consisted four treatments (10 mg kg⁻¹, 100 mg kg⁻¹, 1 000 mg kg⁻¹ and 5 000 mg kg⁻¹) with two replications. The experimental fish was acclimatized in fiber container with 100 L of water for 7 d. Fish with the same weight and size transferred into an aquarium with 5 L of water and stocking density of 10 fishes aquarium⁻¹. The *A. vera* extract added into each aquarium according to the treatments. The survival rate of fish has measured with EPA probit analysis software.

Fish infection

The experimental method used in this study was complete randomized design (CRD) with five treatments and three replications, consisted A (control), B (150 mg kg⁻¹ m), C (300 mg kg⁻¹), D (450 mg kg⁻¹), and E (600 mg kg⁻¹). Infection of fish with *A. hydrophila* bacteria was carried out intramuscularly on fish that had been acclimatized for 7 d. The density of bacteria injected 10⁸ CFU mL⁻¹ as much as 0.1 mL. After being infected, fish reared in an aquarium and given artificial feed twice a day, at 08.00 and 16.00 WIB (West Indonesia Time) as much as 3 % of the body's biomass in an ad libitum. If clinical symptoms occur immediately treated. Clinical symptoms (body damage, food response, shock response) observed after fish were infected with *A. hydrophila* bacteria. If clinical symptoms occur immediately treated with

A. vera extract according to treatment.

Treatment of infected koi fry

Treatment of Koi fry that have been infected with *A. hydrophila* is done by soaking them in a solution of *Aloe vera* extract 48 h. After treatment, the fish were transferred to aquarium maintenance for 14 d while observing their recovery. Observed variables include body damage, response to feeding and behavior ([Haryani et al., 2012](#))

Results and Discussion

In vitro test

The results of the inhibitory zone test (*in vitro*) showed that the concentration of *A. vera* extract at 5 000 mg kg⁻¹ had the largest average inhibition zone of 20.49 mm, whereas at a concentration of 10 mg kg⁻¹ the average smallest inhibition zone was 8.39 mm. *A. vera* concentration of 5 000 mg kg⁻¹ is a relatively strong antibacterial. As the opinion of [Bereksi et al. \(2018\)](#) the diameter of the inhibition zone of 20 mm is categorized as a medium inhibition zone, whereas above 20 mm is categorized as strong. According to [Samsudin et al. \(2018\)](#) the diameter of inhibition zone ≤ 12 mm is categorized as low and the diameter of inhibition zone 13 mm to 14 mm is categorized as medium and the diameter of inhibition zone above 14 cm is categorized as strong in inhibiting bacterial growth. The ability of *A. vera* extract to inhibit the growth of *A. hydrophila* bacteria is due to the presence of secondary metabolite content including polysaccharides (acemaman and glucomannan), flavonoids, tannins and saponins that act as antimicrobials. Glucomannan and acemaman have been proved to accelerate wound healing, activating macrophages, stimulating immune system as well antibacterial and antiviral effects ([Sahu et al., 2013](#)). [Elayaraja et al. \(2015\)](#) stated that antimicrobial activity is caused by the presence of terpenoid compounds in plant extracts that can damage the cell walls of bacteria that form complex compounds with cellular extracts, so that proteins and microbial cell walls are destroyed. Besides that antibacterial activity is also caused by the presence of phenolic compounds (tannins and flavonoids) contained in the extract. [Mandal et al. \(2017\)](#) states the antibacterial activity of plant extracts can be attributed to the high content of phenols and flavonoids. Several studies have shown the antibacterial effect of secondary metabolites in inhibiting various types of bacteria.

Phenolic compounds and especially flavonoids can act as antimicrobial agents through several different mechanisms, including inhibiting nucleic acid synthesis, inhibiting cytoplasmic membrane function, inhibiting energy metabolism, inhibiting biofilm adhesion and formation, inhibiting porin in cell membranes, changing membrane permeability, which can cause damage to membrane cells, and weakening pathogenicity (Safaria and Asbchinb, 2019). Opinion of Sengul *et al.* (2009) phenolic compounds are synthesized by medicinal plants in response to microbial infections, so in vitro testing is an effective antimicrobial agent against a variety of pathogenic microorganisms.

Table 1: Inhibit zone *Aloe vera* extract.

Concentration (mg kg ⁻¹)	Average inhibitory zone
10	8.39
100	9.77
1 000	11.32
5 000	20.49

In vivo test

The results of the LC₅₀ immersion test 48 h (*in vivo*) *A. vera* extract on koi fry showed that the concentration of *A. vera* extract analyzed using EPA Probit Analysis software found a concentration of 1 157.16 mg kg⁻¹ could kill fish as much as 50 % of total individuals. while the concentration of 304.95 mg kg⁻¹ can kill fish as much as 15 % of the total individuals. This shows that the safe concentration for fish for the treatment of disease is below 1 157.16 mg kg⁻¹. Some of the active compound in *A. vera* extract will be toxic if the concentration is too high, especially for saponin. Saponin is completely cytotoxic. This was in line with Septriarusli *et al.* (2012), saponin is a toxin that will ruin the blood cell (hemolysis). The potential of secondary metabolites in plants as antimicrobials can be used by suppressing antimetabolic and toxic contents, one of which is to pay attention to the concentration of the plant extract (Chakraborty, 2013).

Clinical symptoms

Observation of the clinical symptoms of Koi fry infected by *A. hydrophila* includes body damage and changes in the behavior of Koi fry. Body damage includes (ulcer, hemorrhagic, exophthalmia, dropsy, discoloration, ulcers, flaky fins) whereas observed behavioral changes include response to feed and response to shock.

Body damage

Infected Koi fry by *A. hydrophila* shows early clinical symptoms within 7 h after infection. This is in accordance with Haryani *et al.* (2012) research that clinical symptoms due to infection with *A. hydrophila* bacteria in carp fish appear after 4 h to 7 h after infection. The clinical symptoms seen at the outset were inflammation in the injection site (Figure 1a) and red (hemorrhagic) spots in fish (Figure 1b), this occurred in all treatments.

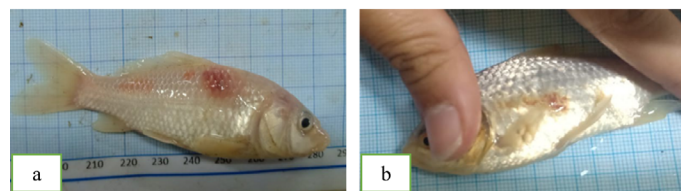


Figure 1: Early clinical symptoms (a) inflammatory, (b) hemorrhagic

According to Haryani *et al.* (2012) clinical symptoms of fish attacked by *A. hydrophila* bacteria or *Motile Aeromonas Septicemia* (MAS) disease characterized by inflammation by swelling and redness of the injection site and then appearing hemorrhagic on the fish body. According to Pandey *et al.* (2010) the emergence of redness (hemorrhagic) on the surface of the fish body is caused by the activity of hemolysin enzymes produced by the bacterium *A. hydrophila* with the target of breaking down red blood cells, so that the cells come out of the blood vessels and cause redness on the skin surface. According to Hamid *et al.* (2016) *A. hydrophila* bacteria degrade body organ tissues and secretes toxins in the form of hemolysin which is spread throughout the body through the bloodstream causing inflammation.

After visible damage to the body in the form of hemorrhagic appeared evenly, Koi fry was immediately given treatment by immersion using aloe vera extract for 48 h. Observation of the clinical symptoms of Koi fry is carried out after infection and treatment with aloe vera extract. After treatment with *A. vera* extract, body damage in Koi fry experienced healing which differed in each treatment and in each time of observation (Table 2).

Based on Table 1 on 1st d to 3rd d post-treatment, koi fry bodies in all treatments experienced ulcers (Figure 2a). According to Lukistiyowati and Kurniasih (2012), ulcers can occur because the regeneration of damaged cells runs slower than the cell death that occurs). This

is reinforced by the statement of [Kozłńska and Pekala \(2012\)](#) that the occurrence of ulcers is caused by the high density of *A. hydrophila* bacteria in the target organs, so that the volume and intensity of toxins released in the infectious process are also higher in that part, while others enter the body blood flow.

Table 2: Damage to Koi fry after treatment.

Day	Body damage				
	A (0 mg kg ⁻¹)	B (150 mg kg ⁻¹)	C (300 mg kg ⁻¹)	D (450 mg kg ⁻¹)	E (600 mg kg ⁻¹)
1	UHP	UHP	UHP	UHGP	UHP
2	UHP	UHP	UHP	UP	-
3	UHPG	UHP	UHP	UP	-
4	UHPG	UPB	UP	UP	-
5	UHP	UPB	UP	H	-
6	UHP	UP	UP	H	-
7	UHPB	P	H	H	-
8	UHPB	P	H	H	-
9	UHPB	P	H	H	-
10	UHPBG	P	H	H	-
11	UHPBGED	H	H	H	-
12	UHPBGE	H	H	H	-
13	UHPG	H	H	H	-
14	UHPG	H	H	H	-

Description: U= Ulcer; B= Enlarged ulcer; H= Hemorrhagic & inflammatory; E= exophthalmia; P= Discolored; D= Dropsy; G= Flake fin; H= Heal; -= Dead.

Koi fry treatment A (control) on the 7th d the ulcer enlarged to become ulcers. According to [Mustaqim et al. \(2017\)](#) ulcers that begin to enlarge can cause the meat to be damaged to cause an enlarged ulcer ([Figure 2b](#)). In addition to experiencing ulcers and enlarged ulcer Koi fry in treatment A also experience prominent eyes (exophthalmia) and distended stomach (dropsy) ([Figure 2c, d](#)). According to [Noor El-Deen et al. \(2013\)](#) exophthalmia occurs because the accumulation of fluid toxins in the eye causes the eyeball to become convex and protrude as a result of bleeding around the eyes, while dropsy occurs because the flow of body fluid stops and seeps out of the capillary and enter into organ, body cavities, and eye cavities. On observation of 1st d of Koi fry in treatment A, body damage was getting worse and worse, resulting in many deaths at the end of the study, because the koi fry in treatment A that are infected with *A. hydrophila* are not given the natural body treatment and defense of the fish which is not able to fight the attack of *A. hydrophila* bacteria.

Koi fry in treatment B (150 mg kg⁻¹) on 4th d and 5th d experienced enlarged ulcers but were not as severe as in treatment A (0 mg kg⁻¹) then fish died on 6th d, ulcers in treated fish B (150 mg kg⁻¹) gradually increased shrinking until the fish only experience pale color on 7th d. According to [Kartikaningsih et al. \(2020\)](#), fish that looks stressed in their body color will look pale and the pattern looks clearer. Koi fry has fully recovered on 11th d, the healing process which is relatively long in treatment B is suspected because antibacterial compounds at a concentration of 150 mg kg⁻¹ are still not enough to treat body damage due to *A. hydrophila* that appears on Koi fry.

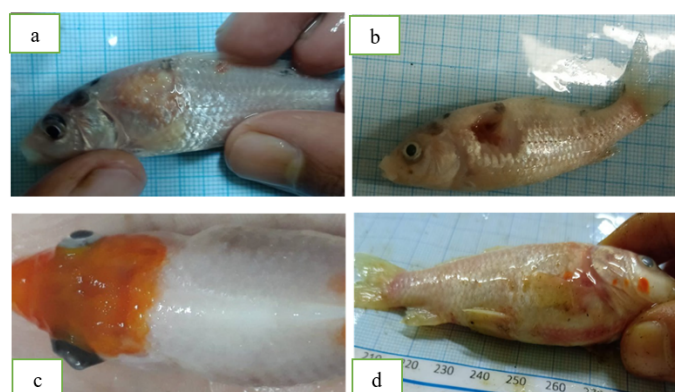


Figure 2: Clinical symptoms of *A. hydrophila* (a) ulcer, (b) enlarged ulcer, (c) exophthalmia, and (d) dropsy

The Koi fry in treatment C (300 mg kg⁻¹) hemorrhagic and ulcers began to heal on the 6th d which was marked by hemorrhagic disappearance and ulcers began to shrink on 4th d ([Figure 3a](#)), total healing occurred on 7th d. Because the concentration of aloe vera extract is high, then the treatment is faster. Seen from healing treatment C (300 mg kg⁻¹) which is faster than treatment B (150 mg kg⁻¹). The higher the concentration of an antibacterial ingredient the stronger the antibacterial activity.

Koi fry in treatment D (450 mg kg⁻¹) on 1st d experienced many deaths, this was caused by the tolerance level of Koi fry to *A. vera* extract only until treatment C (300 mg kg⁻¹) but there were still some fish that survived because of their natural body resistance. against the attack of toxic *A. hydrophila* and saponins. On the 4th day, the Koi fry treatment D that survived experienced faster healing than the other treatments marked by ulcers began to shrink and lasted on the 5th d ([Figure 3a-b](#)).

Koi fry at treatment E (600 mg kg⁻¹) suffered total death on 1st d and 2nd d due to Koi fry which could

not tolerate *A. hydrophila* and saponin compounds produced by *A. vera* extract at a concentration of 600 mg kg⁻¹. Lukistyowati and Kurniasih (2012) stated that the occurrence of fish deaths due to the content of saponins which can cause foam in water that can be toxic to fish. Saponin is a compound that is toxic to fish especially in very dilute solutions and also has hemolysis activity which can damage red blood cells and can result in inhibited breathing processes (Podolak *et al.*, 2010). This shows that the higher the concentration of *A. vera* extract given, the faster the Koi fry recover from MAS disease but decrease at a concentration of 450 mg kg⁻¹ because the saponin content is high so that it is toxic for Koi fry.

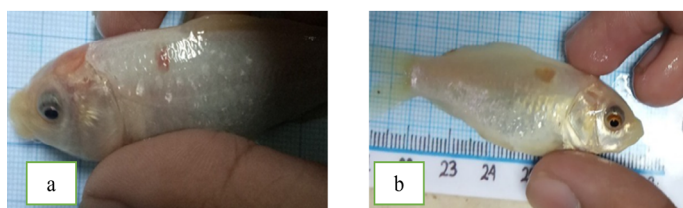


Figure 3: Healing of *A. hydrophila* clinical symptoms (a) ulcers shrink, (b) hemorrhagic loss.

Compared to Tanjung (2014), the healing process of damage to carp body infected with *A. hydrophila* with the treatment of pomegranate peel extract through immersion at a concentration of 100 mg kg⁻¹ tended to be longer ie reaching 11 d to 12 d to recover while the body damage to Koi fry infected with *A. hydrophila* with *A. vera* extract treatment at a concentration of 300 mg kg⁻¹ recovered completely on the 7th d.

The healing process of Koi fry infected with *A. hydrophila* is because *A. vera* extract contains antibacterial compounds, namely flavonoids. Flavonoid compounds are synthesized by plants as a defense system and in response to infections by microorganisms that are effective as antimicrobial compounds against a number of microorganisms (Mierziak *et al.*, 2014). The mechanism of action of anti-inflammatory flavonoids is related to proteins through hydrogen bonds so that the structure of bacterial proteins becomes damaged, cell wall stability and plasma membranes are disrupted, then bacteria undergo lysis, so that the fish gradually heal. In addition to the flavonoids of antibacterial compounds that play an active role in healing are tannins and saponins. Ajizah (2004) explains that the mechanism of the tannin as an antibacterial can kill bacterial growth because the tannin has the power of toxicity which can result in changes in cell permeability. While

saponins also help in the formation of collagen and protein structures that play a role in the process of wound healing (Suratman *et al.*, 1996). Glucomannan also plays a role in the process of wound healing. According to (Mustaqim *et al.*, 2017), glucomannan plays a role in restoring tissue and reducing pain due to disease and protease enzymes in collaboration with glucomannan can break down bacterial cell walls that attack wounds so that it can speed up the wound healing process.

Response to feed

In this current research, as many as 10 primers were occupied for DNA amplification at 15 genotypes of *Phaseolus lunatus* L. The response of eating Koi fry to all treatments after the infection has a tendency to decrease or low due to koi fry stress due to infection with *A. hydrophila*. Hamid *et al.* (2016) stated that the clinical symptoms that caused the change in fish behavior such as swimming on the surface of water, decreased appetite as a result of fish experiencing stress. Olga (2014) stated that the decreasing of feeding response in fish is one of the clinical symptoms of *A. hydrophila* infected the body fish.

Some of the fish in treatment A on the 1st d of observation of the response to feed was low, this continued until the 12th d. It was seen from the amount of leftover food deposited at the bottom of the aquarium when cleaned, then at the end of the observation, there was no response to feed because the fish experienced death (Table 3). Most of the other fish in treatment a had a normal feed response and decreased again on the 13th d. It is suspected that the fish had experienced healing by its natural body resistance until the 13th d, but its body resistance was no longer able to resist the attacks of *A. hydrophila* and Koi fry experiencing metabolic disorders due to the attack of *A. hydrophila*. Roy *et al.* (2018) stated that one of the target organs of attack of *A. hydrophila* is the kidney, where the disruption of these organs can affect the body's metabolic processes.

Treatment B on the 2nd d after the treatment of Koi fry has experienced an increase in feed response until the end of maintenance. Whereas treatment C and D on 3rd d of Koi fry had an increased response to feed but had declined on the 6th d and 7th d but the following day the Koi fry had increased feed response again marked by the depletion of the feed given and the rapid reaction Koi fry for feed given.

Table 3: *The response of Koi fry feed.*

Days to-	Response to Feed														
	A			B			C			D			E		
	Repetition			Repetition			Repetition			Repetition			Repetition		
	1	2	3	1	2	3	1	2	3	1	2	3	1	2	3
1	+	+	+	+	+	+	+	+	+	++	+	+	+	+	-
2	+	+	+	++	++	+	+	++	+	+	++	+	-	-	-
3	+	+	+	++	+	+	++	++	++	++	++	++	-	-	-
4	+	+	+	++	++	++	++	++	++	++	++	++	-	-	-
5	+	+	+	++	++	++	++	++	++	++	++	++	-	-	-
6	+	+	+	++	++	++	++	++	+	++	+	++	-	-	-
7	+	++	++	++	++	++	++	++	+	++	+	++	-	-	-
8	+	++	++	++	++	++	++	++	++	++	++	++	-	-	-
9	+	++	++	++	++	++	++	++	++	++	++	++	-	-	-
10	+	++	++	++	++	++	++	++	++	++	++	++	-	-	-
11	+	++	++	++	++	++	++	++	++	++	++	++	-	-	-
12	+	++	++	++	++	++	++	++	++	++	++	++	-	-	-
13	-	+	+	++	++	++	++	++	++	++	++	++	-	-	-
14	-	+	+	++	++	++	++	++	++	++	++	++	-	-	-

Description: -= No response or dead; += Low response feed; ++= Normal response feed.

Compared with the [Tanjung \(2014\)](#) the normal feed response on carp that were attacked by *A. hydrophila* with the treatment of pomegranate peel extract through immersion took longer, namely on the 7th d while the normal feed response in the Koi fry was attacked by *A. hydrophila* using extract *A. vera* occurs on the 2nd d.

Based on the results of observations, in treatment E on 2nd d, there was no response to feed on Koi fry because the Koi fry had suffered total deaths due to the content of saponin which was toxic at a concentration of 600 mg kg⁻¹.

Treatment by soaking aloe vera extract can restore the feed response of Koi fry attacked by *A. hydrophila* bacteria to normal, while treatment A (control) which was not given aloe vera extract was low until the end of the study. It is assumed that the antibacterial compounds contained in *A. vera* extracts such as flavonoids, tannins, glucomannan, and saponins have reacted and effectively inhibited and damaged v bacterial cells and then repaired damage to the Koi fry organ.

Response to shock

In this current research, as many as 10 primers were

occupied for DNA amplification at 15 genotypes of *P. lunatus*. Response to shock on 1st d in all treatments showed a low response, judging from the low response of fish when tapping the aquarium wall ([Table 4](#)). It is suspected that fish experience post-infection stress and post-immersion. According to [Haryani et al. \(2012\)](#) that Koi fry after infection with *A. hydrophila* bacteria look stressed, swim around aeration and fish swim with sloping body position because the body balance decreases.

Table 4: *Response to Shock Koi Fry.*

Days to-	Response to shock														
	A			B			C			D			E		
	Repetition			Repetition			Repetition			Repetition			Repetition		
	1	2	3	1	2	3	1	2	3	1	2	3	1	2	3
1	+	+	+	+	+	+	+	+	+	++	+	++	+	+	-
2	+	+	+	++	+	+	++	++	++	++	++	++	-	-	-
3	+	++	++	+	++	++	++	++	++	++	++	++	-	-	-
4	+	++	++	++	++	++	++	++	+	++	++	++	-	-	-
5	+	++	++	++	++	++	++	++	+	++	++	++	-	-	-
6	+	++	++	++	++	++	++	++	++	++	++	++	-	-	-
7	+	++	++	++	++	++	++	++	++	++	++	++	-	-	-
8	+	++	++	++	++	++	++	++	++	++	++	++	-	-	-
9	+	++	++	++	++	++	++	++	++	++	++	++	-	-	-
10	+	++	++	++	++	++	++	++	++	++	++	++	-	-	-
11	+	++	++	++	++	++	++	++	++	++	++	++	-	-	-
12	+	++	++	++	++	++	++	++	++	++	++	++	-	-	-
13	-	+	+	++	++	++	++	++	++	++	++	++	-	-	-
14	-	+	+	++	++	++	++	++	++	++	++	++	-	-	-

Description: -= No response or dead; += Low response shock; ++= Normal response shock.

The Koi fry experience changes in behavior after being infected by *A. hydrophila* bacteria, that is, the movement of slow fish and irregular swimming position of fish and always surfaces to look for oxygen. According to [Haryani et al. \(2012\)](#) that goldfish after being infected by *A. hydrophila* bacteria look stressful, swim around clustered around aeration and fish swim with sloping body position because the body balance decreases. [Aminah et al. \(2013\)](#) added that unhealthy or sick fishes look like slow movements of fish, some fish swim close to the surface or around aeration and swim with a sloping body position due to reduced body balance and damage to fish fins due to bacterial attack *A. hydrophila*

The control treatment (A) showed a stagnant or

unchanging response, namely a low response to test 1 until the end of maintenance, but in test 2 and test 3 the shock response became normal and decreased again to a low response to surprises. Treatment B showed a normal shock response on 3rd d to the end, while the shock response of treatment C and D was normal on 6th day, which was marked by Koi fry that had been swimming actively and was not always near aeration. E treatment had no response on 2th d because the fish had experienced total death.

Compared with the Tanjung (2014), the normal shock response in carp attacked by *A. hydrophila* with the treatment of pomegranate peel extract through immersion occurred on the 3rd day while the shock response in the of koi fry was attacked by *A. hydrophila* with the treatment using aloe vera extract gradually normal on the 2nd d. Recovery of a shock response occurs because the antibacterial substances contained in the extract of *A. vera* have entered the body of the fish and are able to damage the bacterial cells of *A. hydrophila*. So that the fish attacked by the disease experience a recovery period and are able to show a normal response. Based on this research comparison of *A. vera* extract research with pomegranate extract research (Tanjung, 2014) the healing process of clinical symptoms including body damage, feed response and shock response on Koi fry attacked by *A. hydrophila* with treatment using *A. vera* extract through immersion is relatively faster compared to treatment using pomegranate peel extract through soaking. Likewise, the research results of Andriani et al. (2020) the healing process of Koi fish infected with *A. hydrophila* using telang (*C. ternatea*) leaf extract was slower (9 d) and lower survival (53.77 %) than using *A. vera* extract. Because *A. vera* extract contains antibacterial compounds such as saponin, acemannan and glucomannan which is not possessed by pomegranate extract (Nozohour et al., 2018)

Conclusions and Recommendations

The results showed that the concentration of 300 mg kg⁻¹ *A. vera* extracts effectively treated the of Koi fry infected with *A. hydrophila* through immersion for 48 h. It can be seen from the healing process of body damage, response to feed and a relatively rapid shock response.

Novelty Statement

Aloe vera extract has the ability as an antibacterial

with inhibition zone diameter reaching 20.49 mm at a concentration of 5 000 mg kg⁻¹ and LC50 test 48 h (*in vivo* test) of 1 157.16 mg kg⁻¹. The investigation of this research found that the concentration of 300 mg kg⁻¹ *Aloe vera* extracts effectively treated the of Koi fry infected with *Aeromonas hydrophila* through immersion for 48 h.

Author's Contribution

RR conceptualized and designed the study, elaborated the intellectual content, performed literature search, data acquisition, data analysis, statistical analysis, manuscript preparation, and manuscript revision. AN, YA and KH performed literature search, carried out experimental studies, data acquisition, data analysis, statistical analysis, and manuscript review. OA and AHP defined intellectual content, literature search, data analysis, manuscript review, manuscript editing, and guarantor. All authors read and approved the final manuscript

Conflict of interest

The authors have declared no conflict of interest.

References

- Ajizah, A. 2004. Sensitivitas *Salmonella typhimurium* terhadap ekstrak daun *Psidium guajava* L. [Sensitivity of *Salmonella typhimurium* to *Psidium guajava* L. leaf extract]. Bioscientiae, 1(1): 31–38. [in Bahasa Indonesia].
- Aminah, S.B. Prayitno and Sarjito. 2014. Pengaruh perendaman ekstrak daun Ketapang (*Terminalia cattapa*) terhadap kelulushidupan dan histologi hati ikan mas (*Cyprinus carpio*) yang diinfeksi bakteri *Aeromonas hydrophila* [The soaking effect of Ketapang leaves (*Terminalia cattapa*) Extract toward survival rate and histological liver of carp infected by *Aeromonas hydrophila*]. J. Aquac. Manage. Technol., 3(4): 118–125. [in Bahasa Indonesia].
- Andriani, Y., A.A. Handaka, Rosidah and R. Himyati. 2020. Potential of Telang plant (*Clitoria ternatea*) for treatment of *Aeromonas hydrophila* infection on Koi Fish (*Cyprinus carpio*). Omni-Akuatika, 16(1): 24–31. <https://doi.org/10.20884/1.oa.2020.16.1.751>
- Bereksi, M.S., H. Hassaine, C. Bekhechi and D.E. Abdelouahid. 2018. Evaluation of antibacterial activity of some medicinal plants extracts

- commonly used in Algerian traditional medicine against some pathogenic bacteria. *Phcog. J.*, 10(3): 507–512. <https://doi.org/10.5530/pj.2018.3.83>
- Chakraborty, S.B., P. Horn and C. Hancz. 2013. Application of phytochemicals as growth-promoters and endocrine modulators in fish culture. *Rev. Aquac.*, 6(5): 1–19. <https://doi.org/10.1111/raq.12021>
- Elayaraja, A., S. Muthupandi, M. Radhakrishnan and S.A. Rahaman. 2015. *In vitro* antioxidant and antibacterial activity of plant extracts of *Pergularia extensa* chiov. *Int. J. Pharmacogn. Phytochem. Res.*, 7(3): 510–512.
- Hamid, N.H., M.D. Hassan, M.Y. Sabri, A.H. Hasliza, R.H. Hamdan and M.N.F. Afifah. 2016. Studies on pathogenicity effect of *Aeromonas hydrophila* infection in juvenile red hybrid tilapia *Oreochromis* sp. *Proc. Int. Semin. Livest. Prod. Vet. Technol., ICARD.* <https://doi.org/10.14334/Proc.Intsem.LPVT-2016-p.532-539>
- Hamouda, T. and J.R. Baker. 2019. Antimicrobial mechanism of action of surfactant lipid preparations in enteric Gram-negative bacilli. *J. Appl. Microbiol.*, 89(3): 397–403. <https://doi.org/10.1046/j.1365-2672.2000.01127.x>
- Haryani, A., R. Grandiosa, I.D. Buwono and A. Santika. 2012. Uji efektivitas daun pepaya (*Carica papaya*) untuk pengobatan infeksi bakteri *Aeromonas hydrophila* pada ikan mas koki (*Carassius auratus*). [Test the effectiveness of Papaya leaves (*Carica papaya*) for the treatment of *Aeromonas hydrophila* bacterial infections in goldfish (*Carassius auratus*)]. *Jurnal Perikanan dan Kelautan.* 3(3): 214–220. [in Bahasa Indonesia].
- Huang, Q., X. Liu, G. Zhao, T. Hu and Y. Wang. 2018. Potential and challenges of tannins as an alternative to in-feed antibiotics for farm animal production. *Anim. Nutr.*, 4(2): 137–150. <https://doi.org/10.1016/j.aninu.2017.09.004>
- Kar, S.K. and T.K. Bera. 2018. Phytochemical constituents of *Aloe vera* and their multifunctional properties: A comprehensive review. *Int. J. Pharm. Sci. Res.*, 9(4): 1416–23.
- Kartikaningsih, H., Yahya, F.Z. Rohman and A.A. Jaziri. 2020. Characteristics of *Aeromonas hydrophila*-infected catfish (*Clarias* sp.). *IOP Conf. Ser. Earth Environ Sci.*, 493(012036): 1–8. <https://doi.org/10.1088/1755-1315/493/1/012036>
- Khan, R.U. and M. Ayub. 2020. Effect of different chemical preservatives on the quality attributes of guava *Aloe vera* blended pulp at ambient conditions. *Sarhad J. Agric.*, 36 (2): 374–733. <https://doi.org/10.17582/journal.sja/2020/36.2.411.418>
- Kozińska, A. and A. Pękala. 2012. Characteristics of disease spectrum in relation to species, serogroups, and adhesion ability of motile aeromonads in fish. *Sci. World J.*, 2012: 1–9. <https://doi.org/10.1100/2012/949358>
- Kraemer, S.A., A. Ramachandran and G.G. Perron. 2019. Antibiotic pollution in the environment: From microbial ecology to public policy. *Microorganisms*, 7(6): 180–185. <https://doi.org/10.3390/microorganisms7060180>
- Kumar, M., G. Gupta, Vikas and S. Sharma. 2018. Feed based vaccine in aquaculture. *Pro. Aqua. Farm. Marine. Biol.*, 1(2): 1–6.
- Laith, A.R. and M. Najiah. 2013. *Aeromonas hydrophila*: Antimicrobial susceptibility and histopathology of isolates from diseased catfish, *Clarias gariepinus* (Burchell). *J. Aquac. Res. Dev.*, 5: 1–7.
- Lukistyowati, I. and Kurniasih. 2012. Pelacakan gen aerolysin dari *Aeromonas hydrophila* pada ikan mas yang diberi pakan ekstrak bawang putih. [Detection of aerolysin gen from *Aeromonas hydrophila* in common carp fed with garlic extract]. *J. Vet.*, 13(1): 43–50. [in Bahasa Indonesia].
- Mandal, S.M., R.O. Dias and O.L. Franco. 2017. Phenolic compounds in antimicrobial therapy. *J. Med. Food*, 20: 1031–1038. <https://doi.org/10.1089/jmf.2017.0017>
- Mierziak, J., K. Kostyn and A. Kulma. 2014. Flavonoids as important molecules of plant interaction with the environment. *Molecules*, 19(10): 16240–16265. <https://doi.org/10.3390/molecules191016240>
- Mustaqim, A., A. Asri and Almurdi. 2017. Pengaruh pemberian gel lidah buaya (*Aloe vera*) terhadap gambaran histopatologi gaster tikus wistar yang diinduksi indometasin. [The effect of *Aloe vera* gel on Wistar rat gastric histopathologic appearance]. *Jurnal Kesehatan Andalas.* 6(3): 641–646. [in Bahasa Indonesia]. <https://doi.org/10.25077/jka.v6i3.751>
- Noor, E-D.A.I., S.I. Shalaby, M.S. Zaki and M.F. Abd Elzاهر. 2013. Some infectious and non-

- infectious eye affection syndrome in fish. Life. Sci. J., 10(2): 1362–1368.
- Nozohour, Y., R. Golmohammadi, R. Mirnejad and M. Fartashvand. 2018. Antibacterial activity of pomegranate (*Punica granatum* L.) seed and peel alcoholic extracts on *Staphylococcus aureus* and *Pseudomonas aeruginosa* isolated from health centers. J. Appl. Biotechnol., 5(1): 32–36. <https://doi.org/10.29252/JABR.01.01.06>
- Olga, O., 2014. Patogenisitas bakteri *Aeromonas hydrophila* ASB01 pada ikan Gabus (*Ophicephalus striatus*). [Pathogenicity of *Aeromonas hydrophila* ASB01 bacteria in Cork fish (*Ophicephalus striatus*)]. Sains Akuatik, 14(1): 33–39. [in Bahasa Indonesia].
- Panche A.N., A.D. Diwan, and S.R. Chandra. 2017. Flavonoids: An overview. J. Nutr. Sci., 5: 1–15. <https://doi.org/10.1017/jns.2016.41>
- Pandey, A., M. Naik and S.K. Dubey. 2010. Hemolysin, protease, and EPS producing pathogenic *Aeromonas hydrophila* strain An4 shows antibacterial activity against marine bacterial fish pathogens. J. Mar. Res., 2010: 1–9. <https://doi.org/10.1155/2010/563205>
- Peatman, E., H. Mohammed, A. Kirby, C.A. Shoemaker, M.A. Yildirim and B.H. Beck. 2018. Mechanisms of pathogen virulence and host susceptibility in virulent *Aeromonas hydrophila* infections of channel catfish (*Ictalurus punctatus*). Aquaculture, 482: 1–8. <https://doi.org/10.1016/j.aquaculture.2017.09.019>
- Podolak, I., A. Galanty and D. Sobolewska. 2010. Saponins as cytotoxic agents: A review. Phytochem. Rev. 9(3): 425–474. <https://doi.org/10.1007/s11101-010-9183-z>
- Quezada, M.P., C. Salinas, M. Gotteland and L. Cardemil. 2017. Acemannan and fructans from *Aloe vera* (*Aloe barbadensis* Miller) plants as novel prebiotics. J. Agric. Food. Chem., 65: 10029–10039. <https://doi.org/10.1021/acs.jafc.7b04100>
- Roy, A., J. Singha and T.J. Abraham. 2018. Histopathology of *Aeromonas caviae* infection in challenged Nile tilapia *Oreochromis niloticus* (Linnaeus, 1758). Int. J. Fish. Aquac., 8(20): 151–155. <https://doi.org/10.5376/ija.2018.08.0020>
- Safaria, M. and A.S. Asbchinb. 2019. Evaluation of antioxidant and antibacterial activities of methanolic extract of medlar (*Mespilus germanica* L.) leaves. Biotechnol. Biotechnol. Equip., 33(1): 372–378. <https://doi.org/10.1080/13102818.2019.1577701>
- Sahu, P.K., D.D. Giri, R. Singh, P. Pandey, S. Gupta, A.K. Shrivastava, A. Kumar and D.K. Pandey. 2013. Therapeutic and medicinal uses of *Aloe vera*: A review. Pharm. Pharmacol., 4(8): 1–13.
- Samsudin, N.I.P., H.Y. Lee, P.E. Chern, C.T. Ng, L. Panneerselvam, S.Y. Phang, W.T. Tan and N.A. Mahyudin. 2018. *In vitro* antibacterial activity of crude medicinal plant extracts against ampicillin+ penicillin-resistant *Staphylococcus aureus*. Int. Food Res. J., 25(2): 573–579.
- Sengul, M., H. Yildiz, N. Gungor, B. Cetin, Z. Eser and S. Ercisli. 2009. Total phenolic content, antioxidant and antimicrobial activities of some medicinal plants. Pak. J. Pharm. Sci., 22: 102–106.
- Septiarusli, I.E., H. Kiki, M. Yenny and M. Danar. 2012. Potential secondary metabolite compounds from seed extract of Keben fruit *Barringtonia asiatica* in anesthesia process of tiger grouper *Ephinephelus fuscoguttatus*. J. Fish. Mar., 3: 295–299.
- Sopiah, S., R. Rosidah, W. Lili, I. Iskandar and I.B. Suryadi. 2018. The effectivity of pandanus leaf extract for the treatment of Sangkuriang catfish juvenile *Clarias gariepinus* infected by *Aeromonas hydrophila*. J. Akuakultur Indonesia, 17(2): 120–129.
- Suratman, S.A. Sumiwi and D. Gozali. 1996. Pengaruh ekstrak antanan dalam bentuk salep, krim dan jelly terhadap penyembuhan luka bakar. [Effects of Antananas extract in the form of ointments, creams and jelly on healing burns]. Cermin Dunia Kedokteran, 108: 31–36. [in Bahasa Indonesia].
- Tanjung, A.F., 2014. Efektivitas ekstrak kulit buah delima dalam pengobatan benih ikan mas (*Cyprinus carpio*) yang terinfeksi bakteri *Aeromonas hydrophila*. [Effectiveness of pomegranate skin extract in the treatment of carp (*Cyprinus carpio*) seeds infected by *Aeromonas hydrophila* bacteria]. [Undergraduate Thesis]. Faculty of Fisheries and Marine Science, Padjadjaran University, Bandung. [in Bahasa Indonesia].
- Triyanti, R. and M. Yulisti. 2012. Rantai pemasaran ikan koi (*Cyprinus carpio*) di Kabupaten Blitar, Jawa Timur. [Marketing chain of koi fish (*Cyprinus carpio*) in Blitar Regency, East Java]. Buletin Riset Sosek Kelautan dan Perikanan, 7(1): 14–20. [in Bahasa Indonesia]. <https://doi.org/10.1080/13102818.2019.1577701>

[org/10.15578/marina.v7i1.4593](https://doi.org/10.15578/marina.v7i1.4593)

United State Environmental Protection Agency (USEPA). 2002. Methods for measuring the acute toxicity of effluents and receiving waters to freshwater and marine organisms. Office of Water (4303T) 1200 Pennsylvania Avenue, NW Washington, DC 20460.

Wassif, I.M., 2018. Biochemical and molecular characterization of *Aeromonas* species isolated from fish. *Alex. J. Vet. Sci.*, 57 (1): 38–45.
<https://doi.org/10.5455/ajvs.293293>